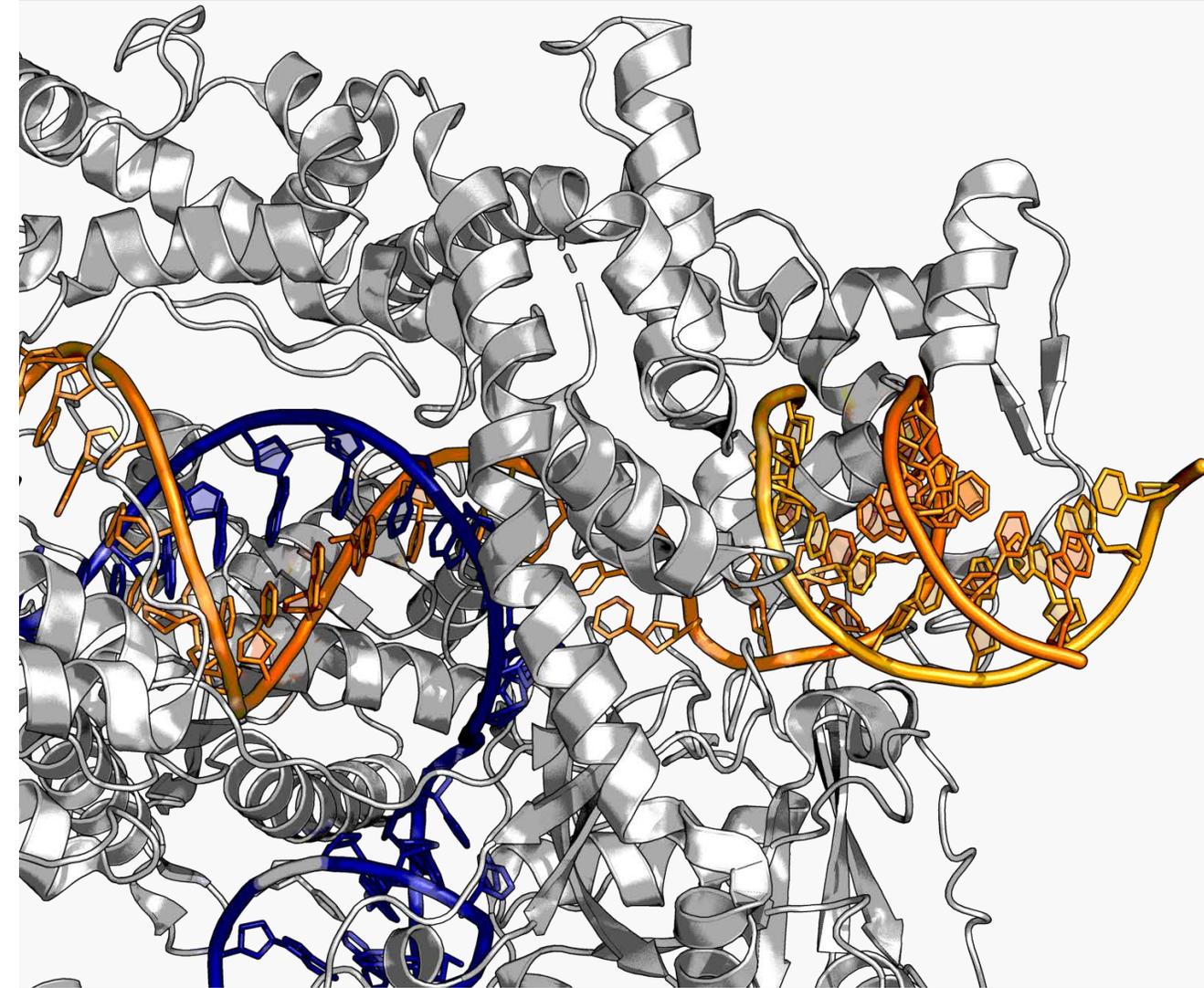


# Enhancing Genome Editing with Engineered CRISPR Enzymes



CENTER OF  
GENOMIC  
MEDICINE



MASSACHUSETTS  
GENERAL HOSPITAL



HARVARD  
MEDICAL SCHOOL

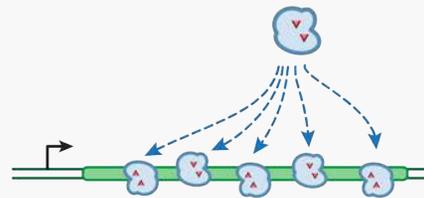
Sunday April 28<sup>th</sup>, 2019

# Outline: Expanding the genome editing toolbox

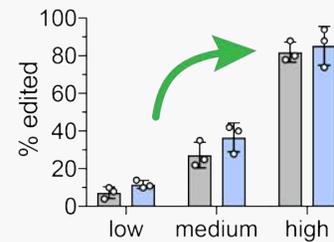


## Introduction: Genome editing and targeting range

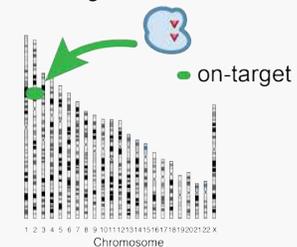
### Part 1: Engineering improved CRISPR-Cas12a enzymes



targeting range

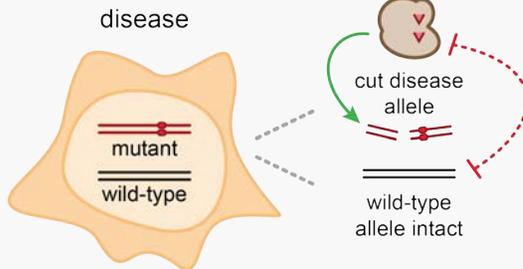


activity



specificity

### Part 2: Challenges & applications



- assay development
- protein engineering
- molecular medicines



Russell



Katie



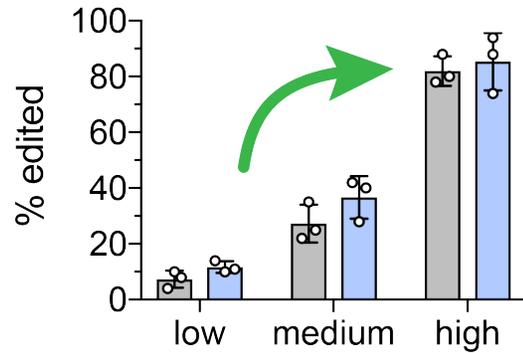
Joey

Kleinstiver Lab

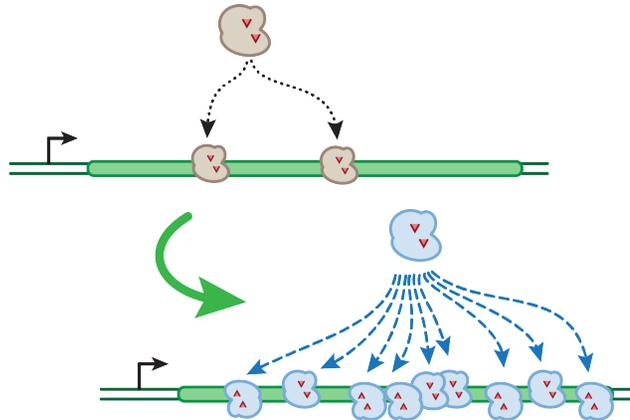
# Ideal genome editing properties



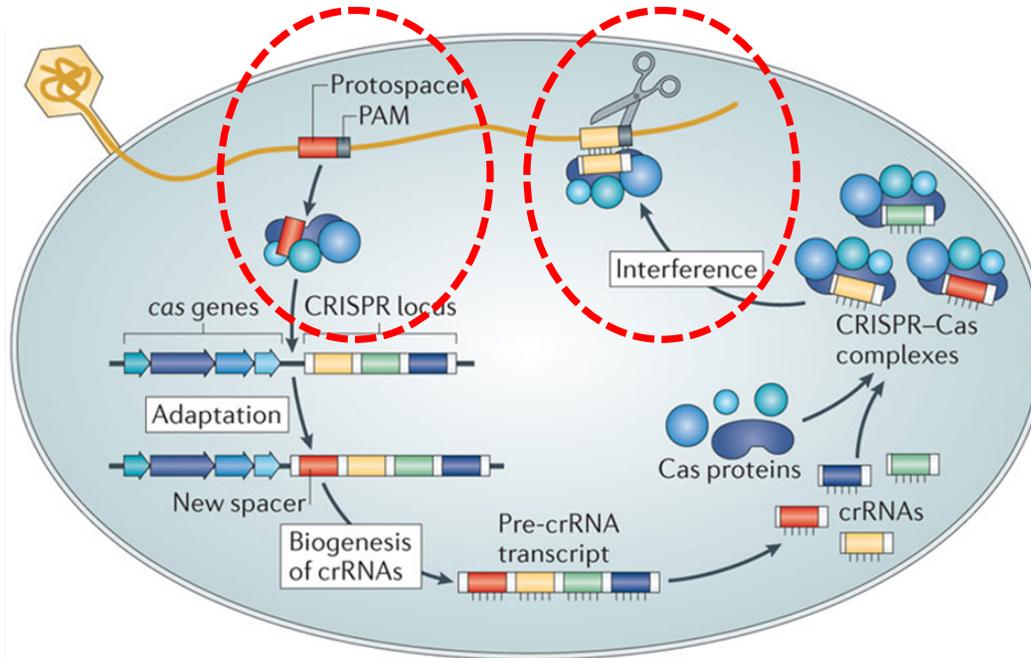
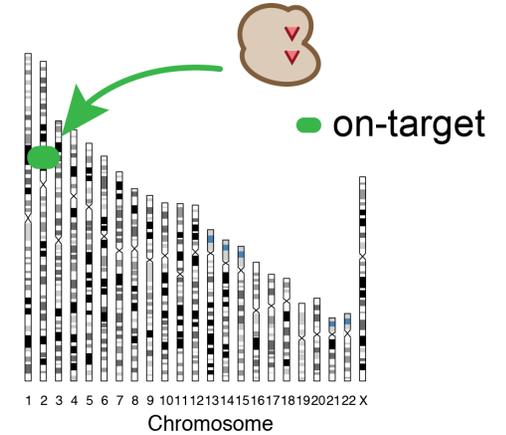
High activity



Broadly targetable



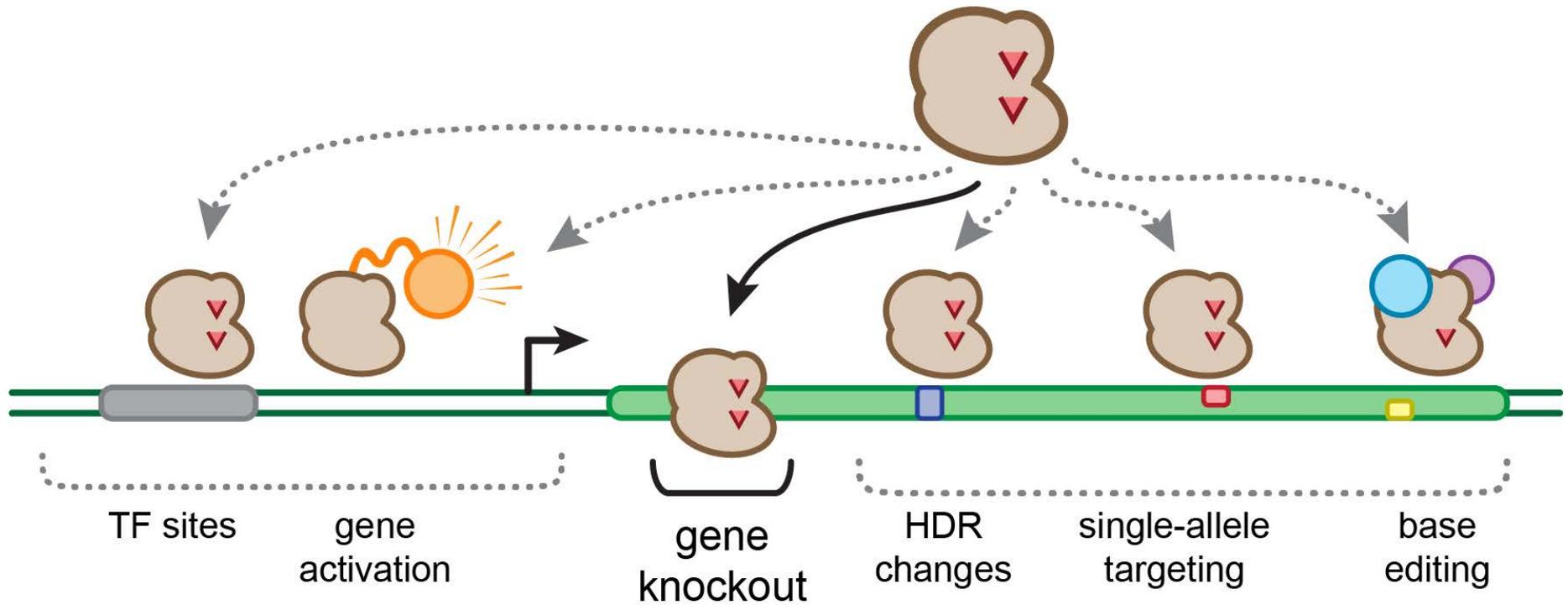
Safe / specific



**Protein engineering**  
to impart desirable  
properties



# PAM preference can limit editing applications



$$\text{SpCas9 PAM} = \text{NGG} = \frac{1}{8} \text{ bp}$$



$$\text{SaCas9 PAM} = \text{NNGRRT} = \frac{1}{32} \text{ bp}$$

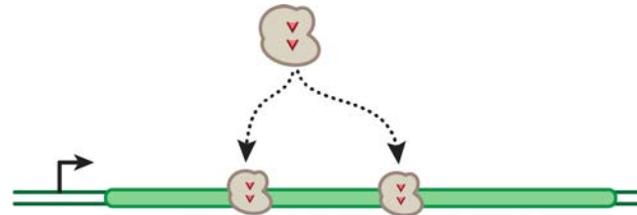


$$\text{Cas12a PAM} = \text{TTTV} = \frac{1}{43} \text{ bp}$$

# Ways to expand targeting via protein engineering



wild-type SpCas9 = NG**G**



[ 1 ]  
alter PAM

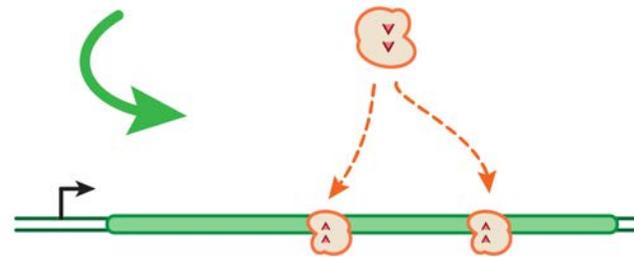
**SELECTIVE**

SpCas9-VRQR = NG**A**

Kleinstiver et al., *Nature*, 2016

SpCas9-VRER = NG**CG**

Kleinstiver et al., *Nature*, 2015

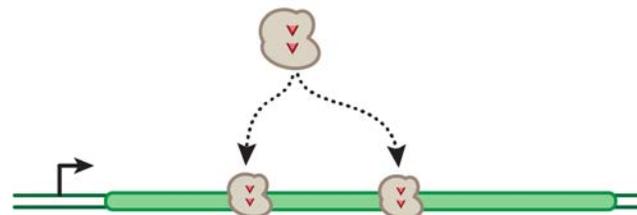


xCas9 = NG**N**

Hu et al., *Nature*, 2018

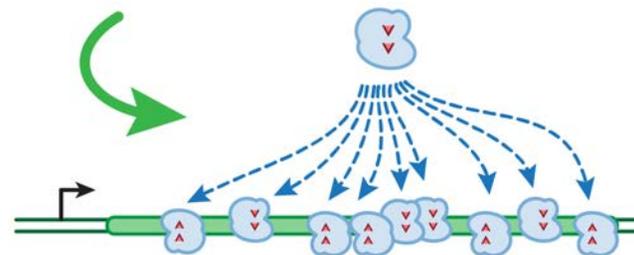
SpCas9-NG = NG**N**

Nishimasu et al., *Science*, 2018



[ 2 ]  
relax PAM

**EXPANDED**

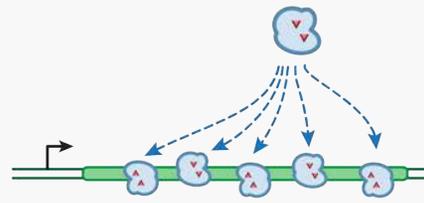


# Outline: Expanding the genome editing toolbox

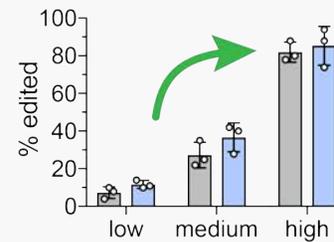


**Introduction:** Genome editing technologies

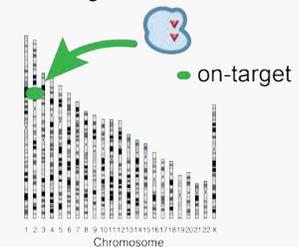
## Part 1: Engineering improved CRISPR-Cas12a enzymes



targeting range



activity



specificity

## Part 2: Challenges & applications



- assay development
- protein engineering
- molecular medicines



Russell



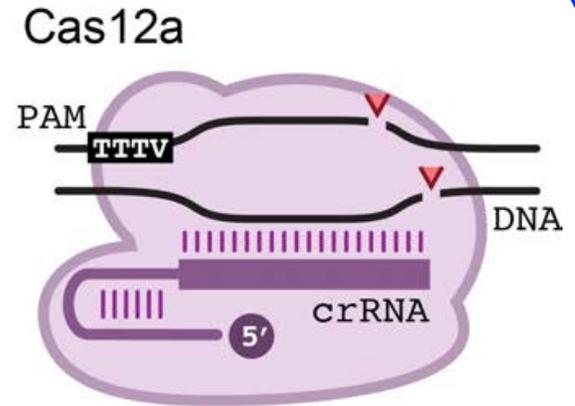
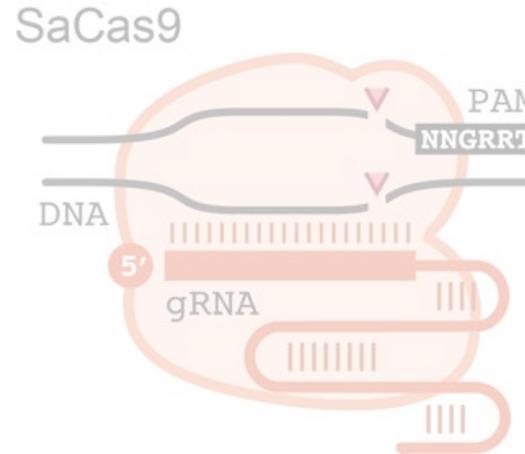
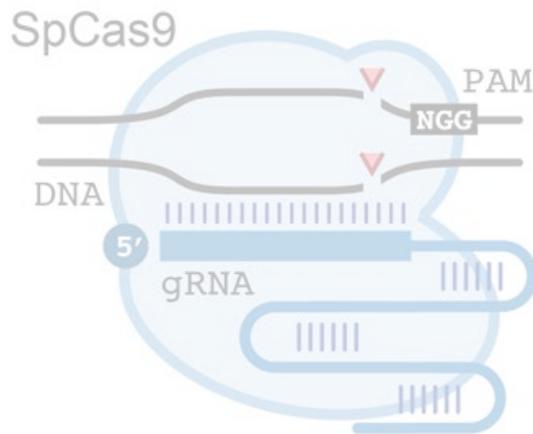
Katie



Joey

Kleinstiver Lab

# PAM requirement prohibits wide use of Cas12a



**CRISPR type**  
**Size (# AA)**  
**PAM**  
**DNA break**  
**guide RNA**  
**multiplex**

type II-A  
 1368  
 NGG (3')  
 blunt, PAM prox.  
 ~100nt (sgRNA)  
 challenging



type II-A  
 1053  
 NNGRRT (3')  
 blunt, PAM prox.  
 ~100nt (sgRNA)  
 challenging



type V-A  
 ~1300  
 TTTV (5')  
 5' overhang, PAM distal  
 ~40nt, single RNA  
 simple

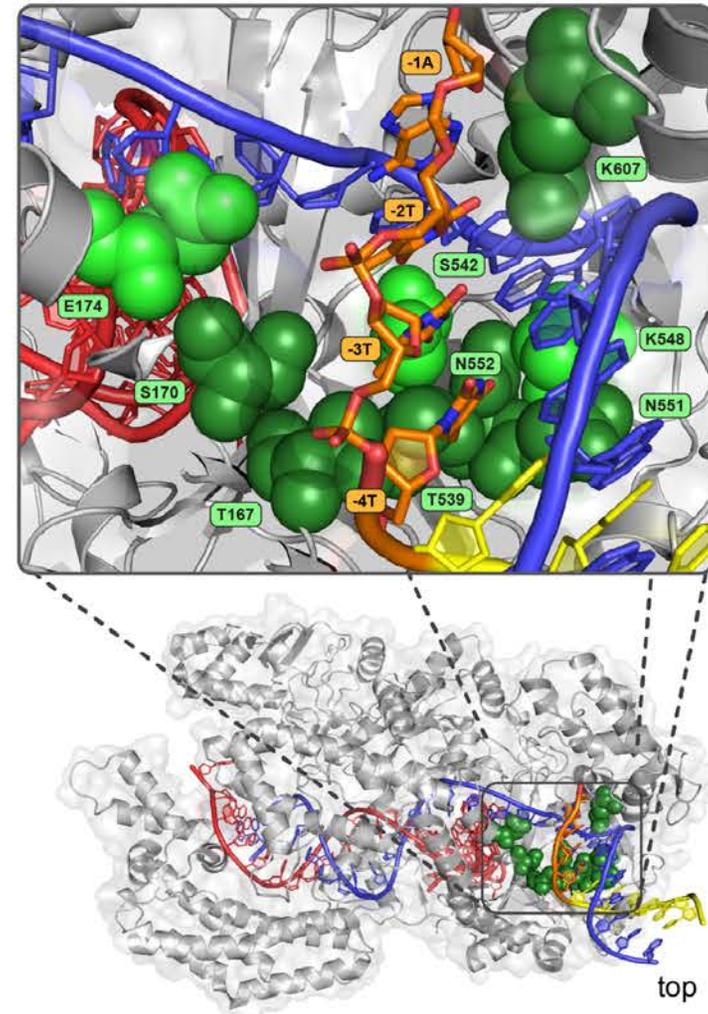
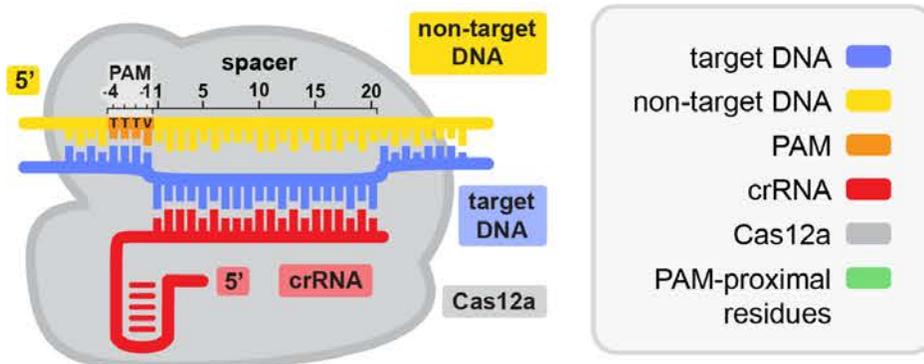


# Improving CRISPR-Cas12a targeting range



Methods to improve properties:

- a. Orthologs
- b. **Engineering**
  - i. Directed evolution
  - ii. **Structure-guided**



**enAsCas12** → expanded PAM preference

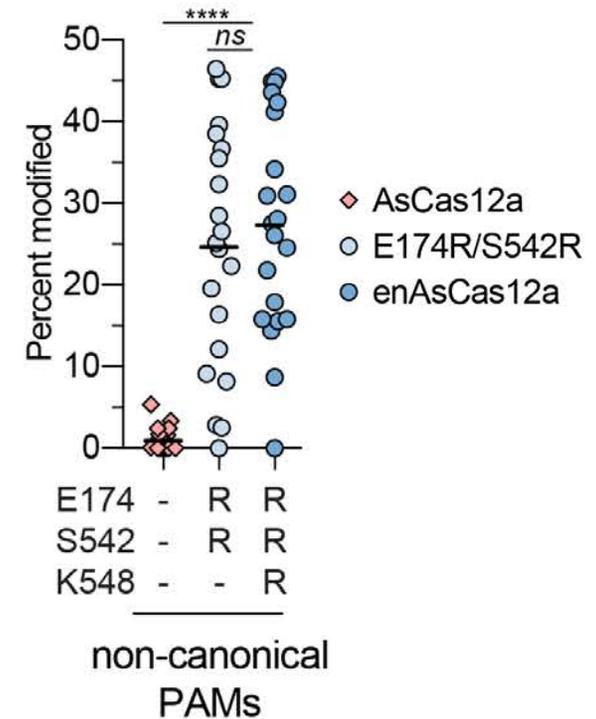
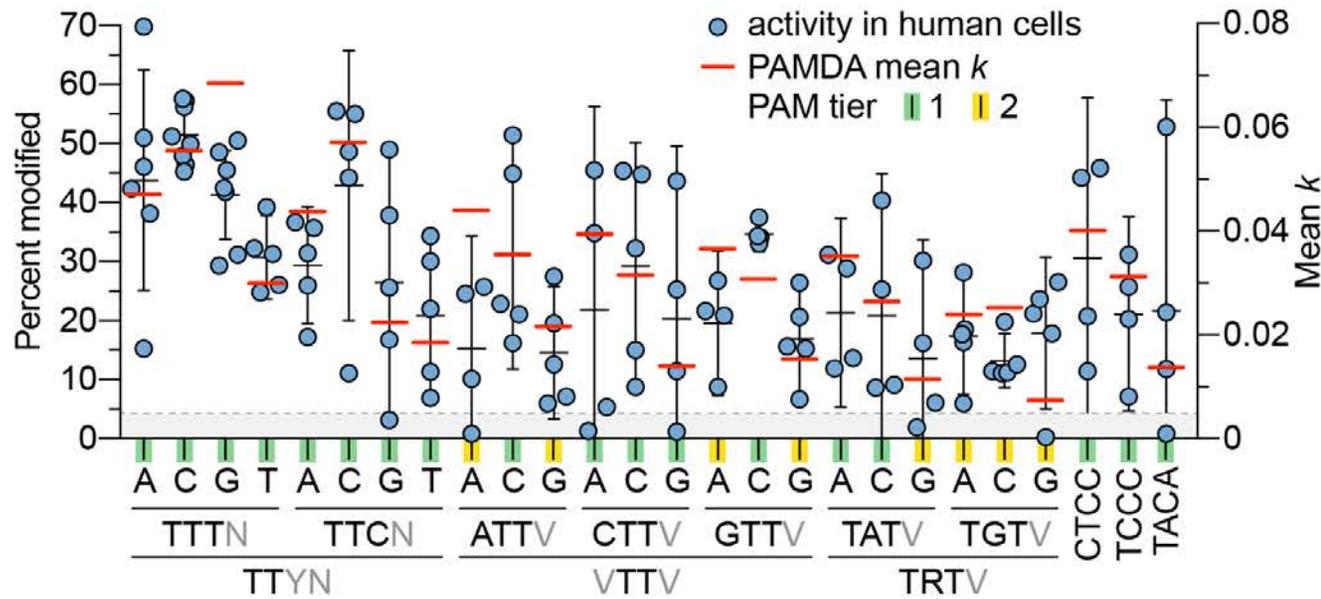


Alex



Russell

# Expanded targeting in human cells with enAsCas12a



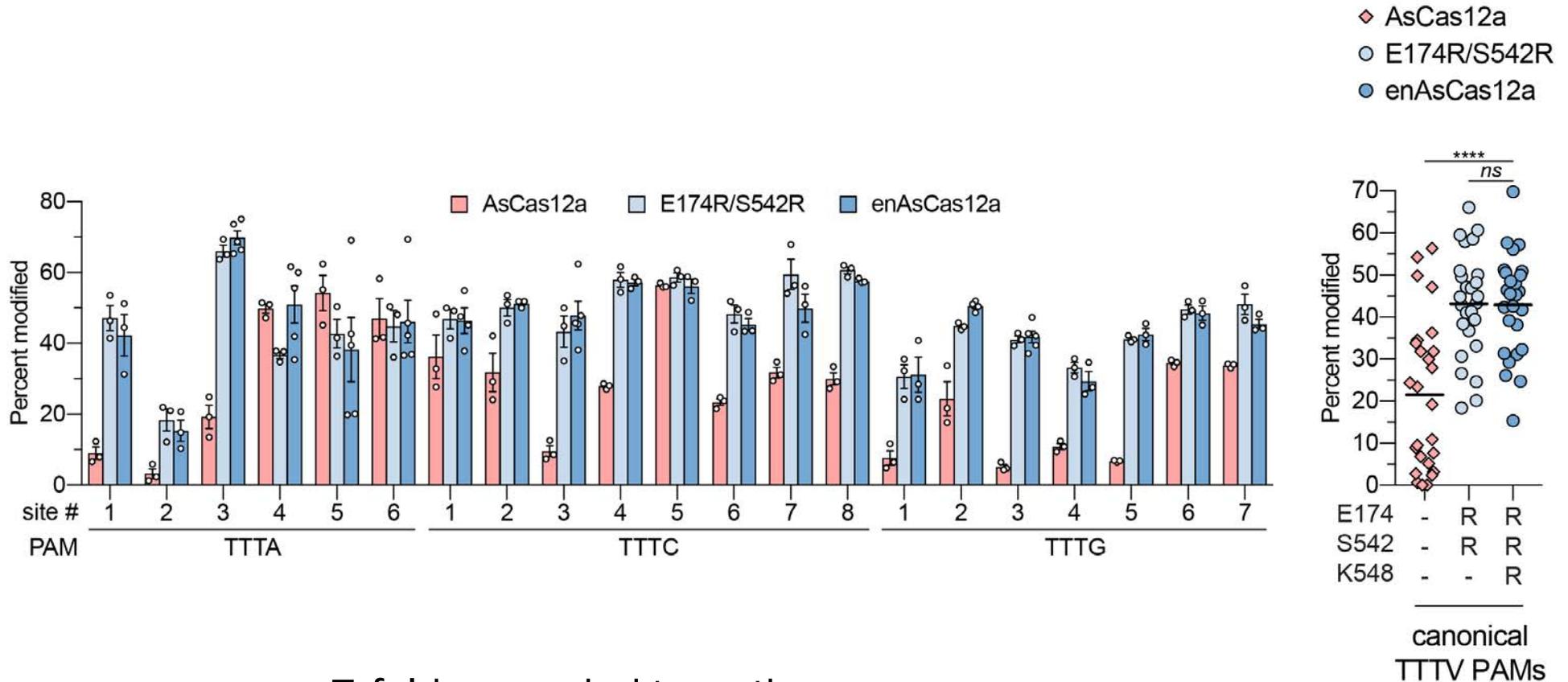
AsCas12a = **TTTV**

enAsCas12a = **TTTV**, TTTT, TTCN, VTTV, TRTV, and more

**~7-fold expanded targeting range**

**[ 2 ] relax PAM preference**

# Improved nuclease activity with enAsCas12a

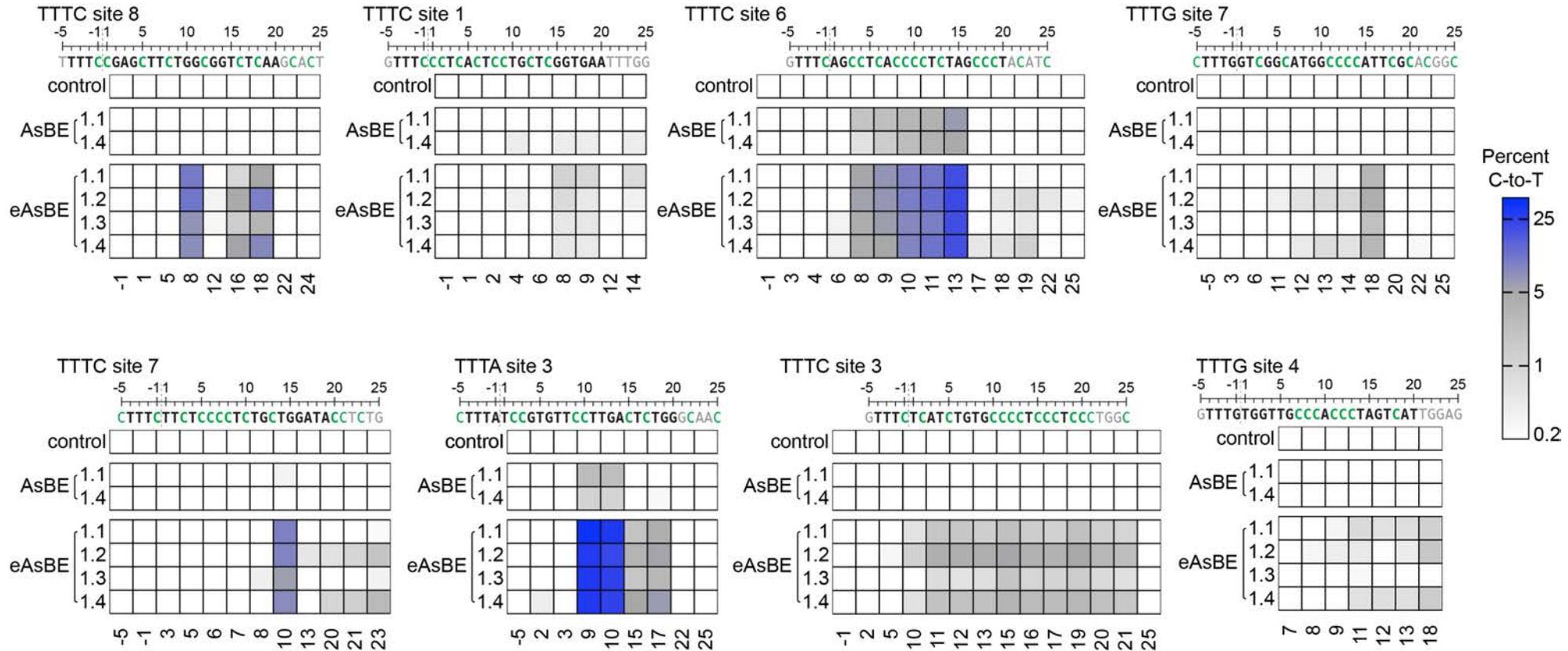


~7-fold expanded targeting range  
**~2-3 fold improved on-target activity**



enAsCas12a

# Improved base editing with enAsCas12a



enAsCas12a

~5-10 fold improved base editor activity

# Summary 1: Improved activities of enAsCas12a



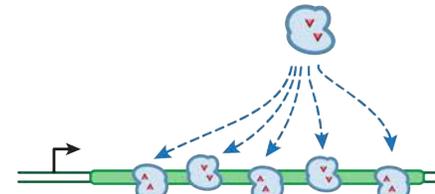
enAsCas12a



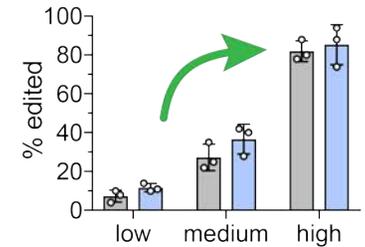
Engineered CRISPR-Cas12a variants with increased activities and improved targeting ranges for gene, epigenetic and base editing

Benjamin P. Kleinstiver<sup>1,2,3,4,10\*</sup>, Alexander A. Sousa<sup>1,2,3,12</sup>, Russell T. Walton<sup>1,2,3,10,12</sup>, Y. Esther Tak<sup>1,2,3,4</sup>, Jonathan Y. Hsu<sup>1,2,3,5</sup>, Kendell Clement<sup>1,2,4,6</sup>, Moira M. Welch<sup>1,2,3</sup>, Joy E. Horng<sup>1,2,3</sup>, Jose Malagon-Lopez<sup>1,2,3,4,7,11</sup>, Irene Scarfò<sup>2,8,9</sup>, Marcela V. Maus<sup>2,8,9</sup>, Luca Pinello<sup>1,2,4,6</sup>, Martin J. Aryee<sup>1,2,4,6,7</sup> and J. Keith Joung<sup>1,2,3,4\*</sup>

Kleinstiver et al., *Nature Biotechnology*, 2019

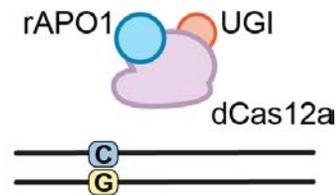


targeting range

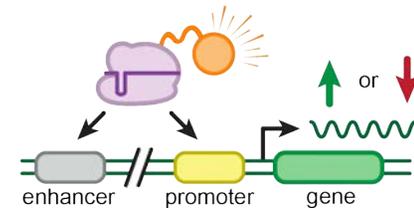


activity

- ~7-fold **expanded** targeting range
- ~2-3 fold improved on-target activity
- enhanced activities at lower temperatures
- can improve other variants (~2-fold, enRVR & enRR)
- improved base editing & gene activation
- efficient editing in primary human T cells



base editing



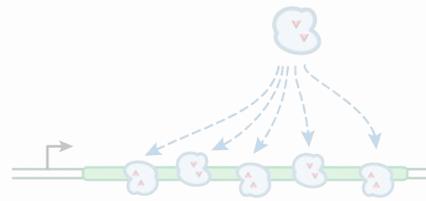
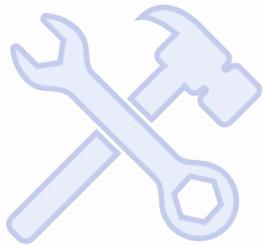
epigenome editing

# Outline: Expanding the genome editing toolbox

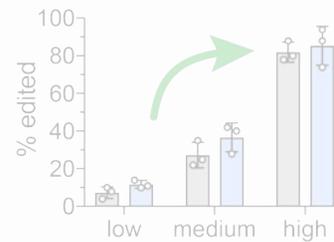


**Introduction:** Genome editing technologies

**Part 1:** Engineering improved CRISPR-Cas12a enzymes



targeting range

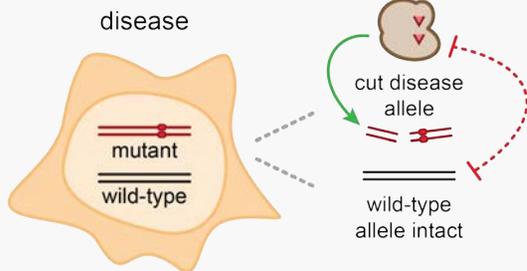


activity



specificity

**Part 2:** Challenges & applications



- assay development
- protein engineering
- molecular medicines



Russell



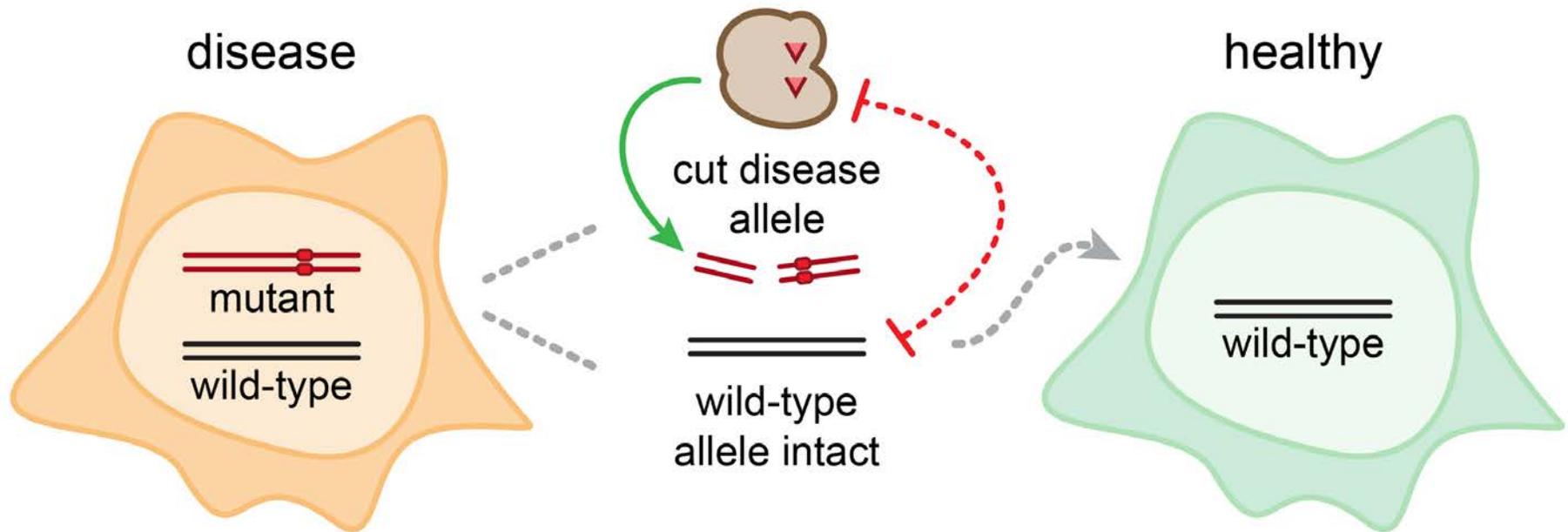
Katie



Joey

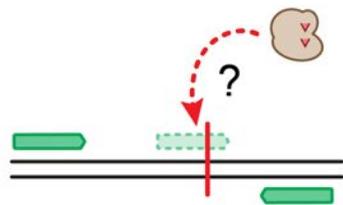
Kleinstiver Lab

# Challenge: Single allele editing



## Important properties for allele-specific editing

targeting range



Position the SNP in PAM

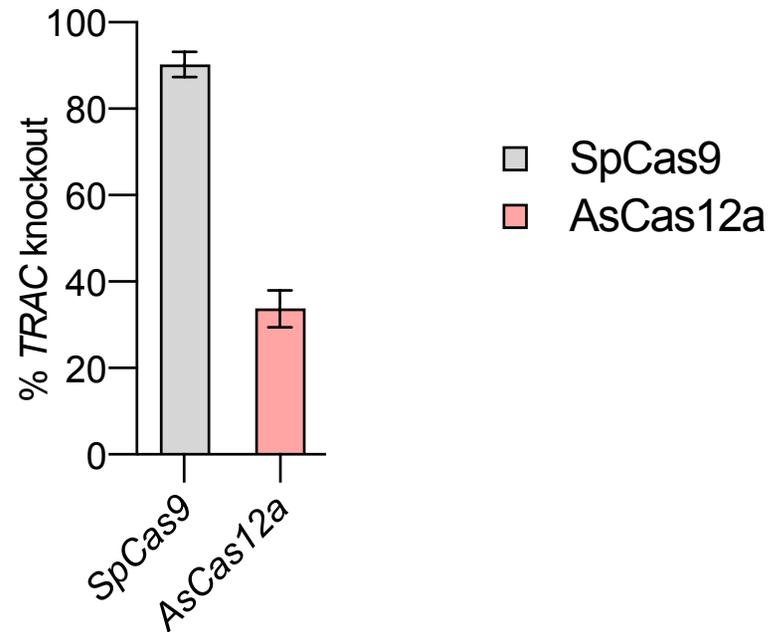
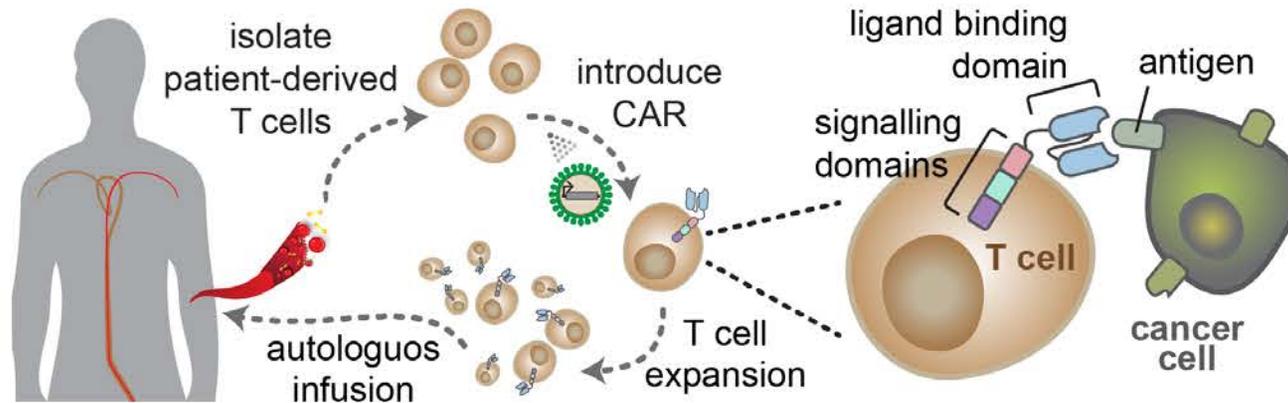
NGG

**Mut** - GATCCAATCGGAATCGGATTTCG

**WT** - GATCCAATCGGAATCGTATTTCG

Need library of PAM selective variants

# Challenge: Assessing improved tools in primary cells



# Building a primary human T cell editing workflow



## (1) T cells



Leukopak

1. RosetteSep Human T Cell Enrichment Cocktail

2. Ficoll-Paque Plus



Isolated T cells

## (2) CRISPR nucleases



SpCas9

+



sgRNA



SpCas9 RNP



AsCas12a

+



crRNA



AsCas12a RNP

## (3) T cell editing workflow



thaw T cells

add  
IL-2 + PHA

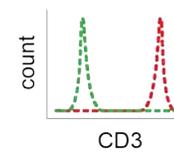
2 or 4 days

electroporate  
*Lonza 4D*

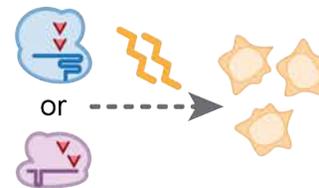
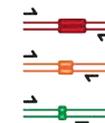
3 days

assess gene editing

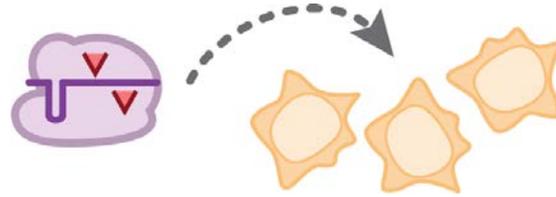
1. staining



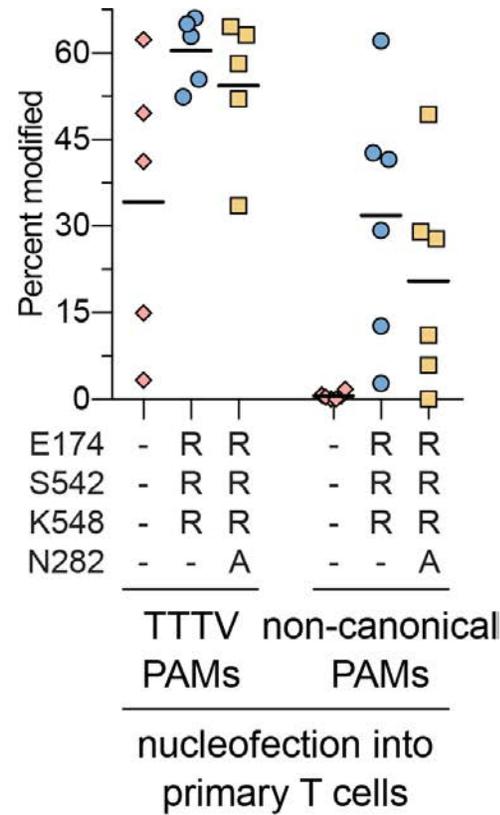
2. genetic analysis



# RNP delivery of enAsCas12a in human cells

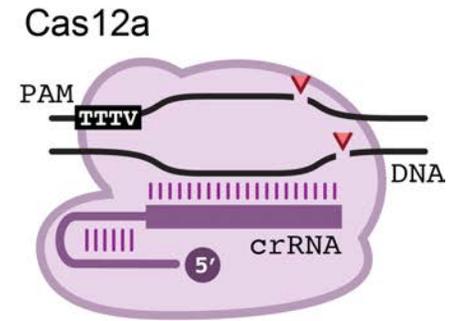
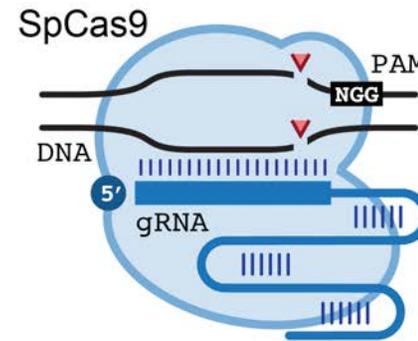
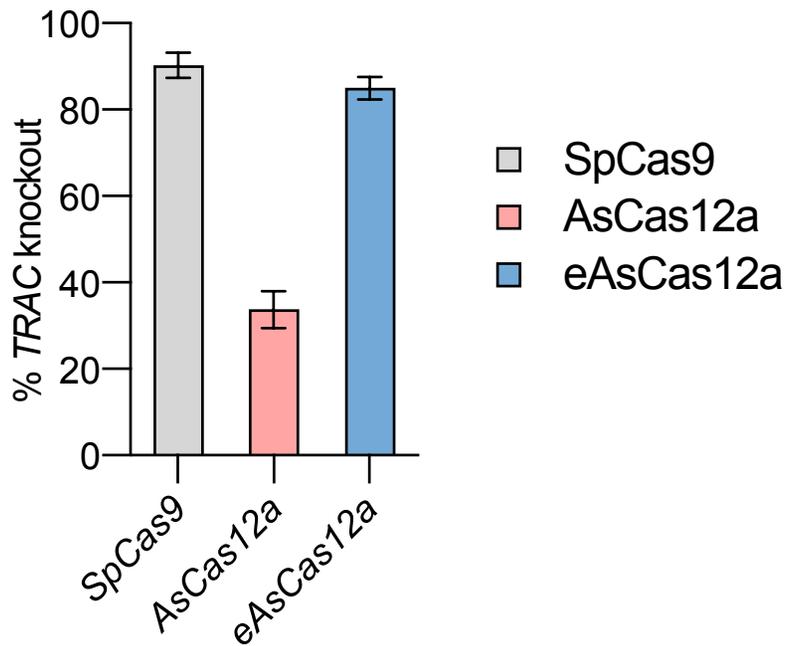


- ◆ AsCas12a
- enAsCas12a
- enAsCas12a-HF1

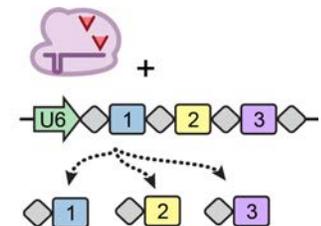
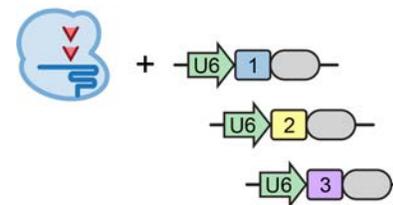


Cas12a

# RNP delivery of CRISPR nucleases in primary T cells



CRISPR type	type II-A	type V-A
Size (# AA)	1368	~1300
PAM	NGG (3')	TTTV (5')
DNA break	blunt, PAM prox.	5' overhang, PAM distal
guide RNA	~100nt (sgRNA)	~40nt, single RNA
multiplex	challenging	simple

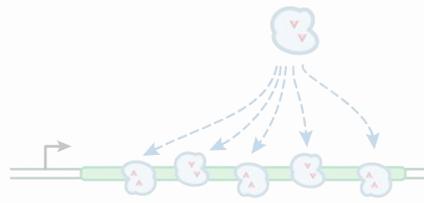
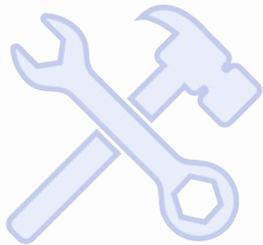


# Summary 2: Addressing challenges of genome editing

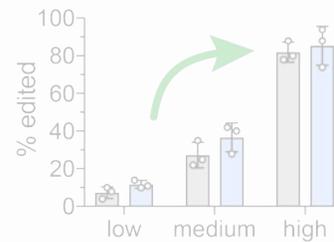


**Introduction:** Genome editing technologies

**Part 1:** Engineering improved CRISPR-Cas12a enzymes



targeting range

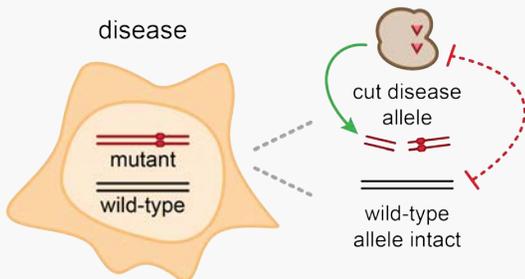


activity



specificity

**Part 2:** Challenges & applications



- assay development
- **PAM selective** Cas9 variants
- Activities in 1<sup>o</sup> T cells



Russell



Katie



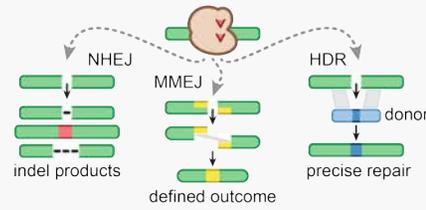
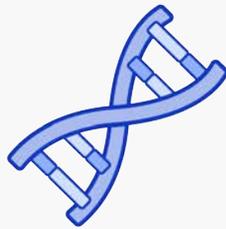
Joey

Kleinstiver Lab

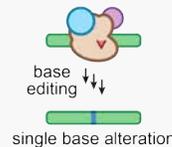
# Summary + Research in the Kleinstiver lab



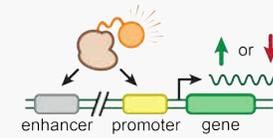
## Genome editing technologies



genome editing

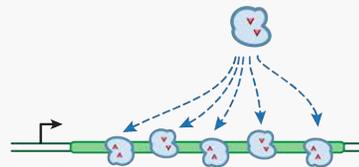


base editing

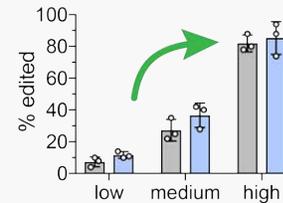


epigenome editing

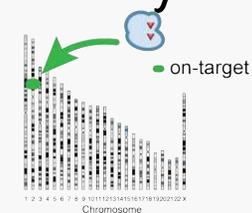
## Protein engineering to enhance CRISPR enzymes



targeting range

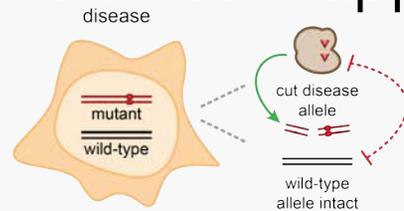


activity

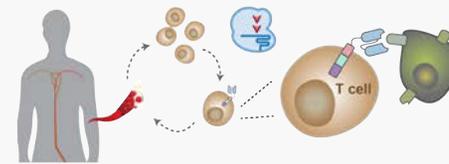


specificity

## Molecular medicines + applications



allele specificity



disease applications

**Thanks** to those who help ask & answer questions



## Joung Lab

Alexander Sousa

Russell Walton

Moira Welch

Esther Tak

Joy Horng

Jon Hsu, et al.

**Martin Aryee** – MGH

Jose Lopez, Sara Garcia

**Luca Pinello** – MGH

Kendell Clement

**Marcela Maus** - MGH

Irene Scarfo



[www.kleinstiverlab.org](http://www.kleinstiverlab.org)



Colleagues & Collaborators  
Friends & Family

## Kleinstiver Lab



Russell  
Walton



Katie  
Christie



Joey  
Rissman

*Recruiting postdocs,  
students, & technicians*

