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## Presenter Disclosure Ryan L. Boudreau, Ph.D

**The following relationships exist related to this presentation:**

**No relationships to disclose.**

2010 ASGCT Annual Meeting, Session 121: Emerging Field Review: Regulatory RNAs

19 May 2010

# RNAi-based Strategies for Silencing Disease

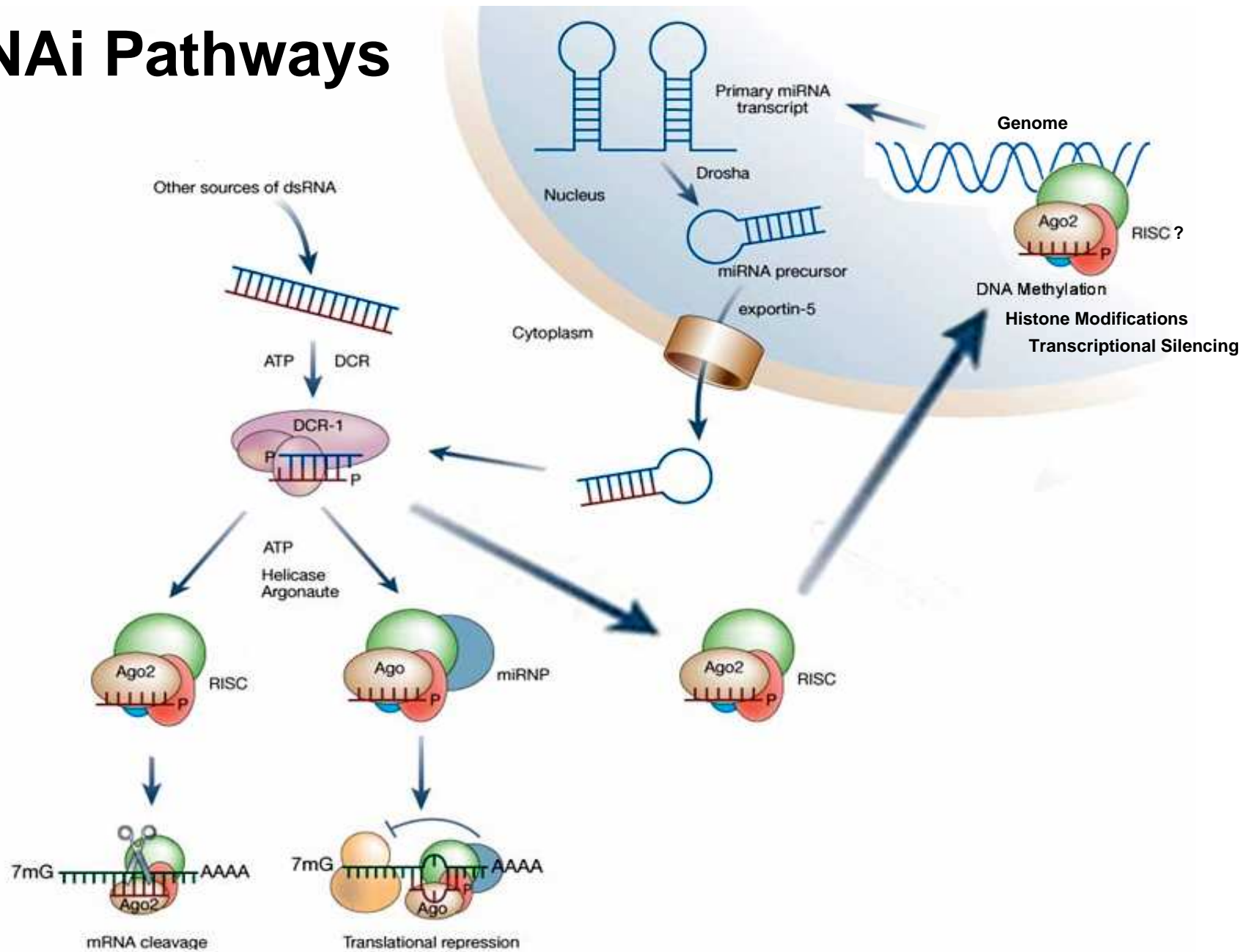
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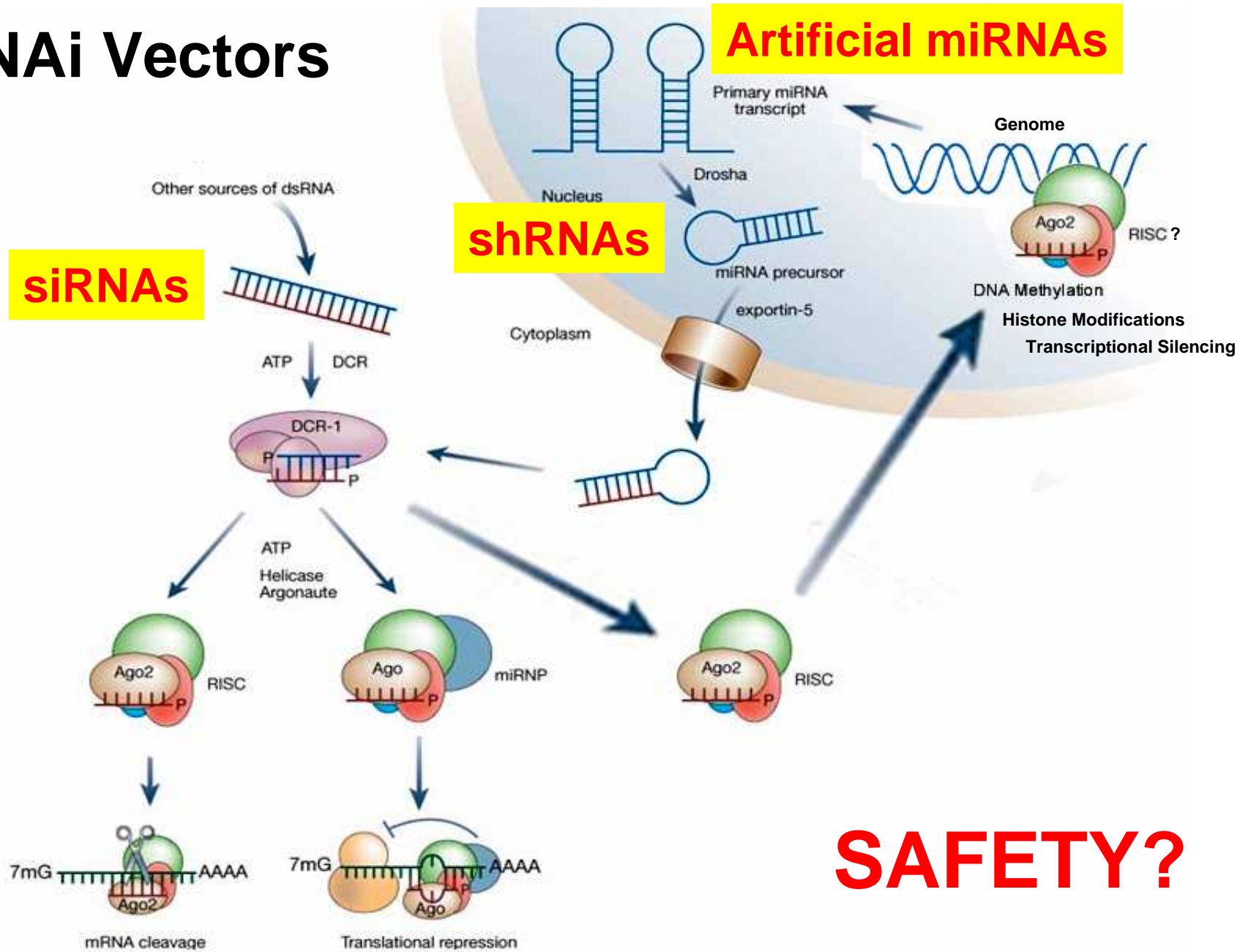
# RNA Interference (RNAi)

- Process of gene regulation mediated by small dsRNAs
- Utilized as a common biological tool to study gene function
- Under investigation as a therapeutic to treat various diseases for which silencing a target may provide benefit
  - Viral infection (e.g. HIV, RSV, HBV and SARS among others)
  - Cancer (e.g. glioma, melanoma or targets common to most cancers)
  - Genetic diseases (e.g. Huntington's, Alzheimers, ataxias, etc.)
- RNAi Delivery or Expression
  - Exogenous synthetic siRNAs (transient gene silencing)
  - Vectors expressing stem-loop RNAs (persistent gene silencing)
- Absolute specificity is questionable
  - Cellular responses to dsRNA (IFN- $\beta$ , PKR, OAS1)
  - Off-target effects due to saturation of RNAi machinery or via partial complementarity with unintended mRNAs
  - Supports the need for optimizing RNAi vectors and potentially developing tissue-specific and regulated expression strategies

# RNAi Pathways



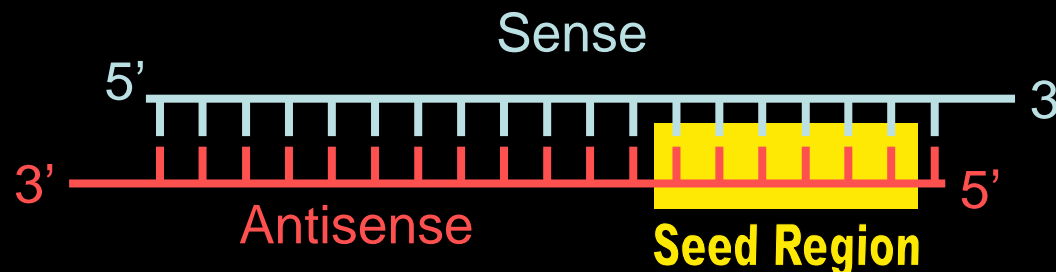
# RNAi Vectors



# SAFETY?

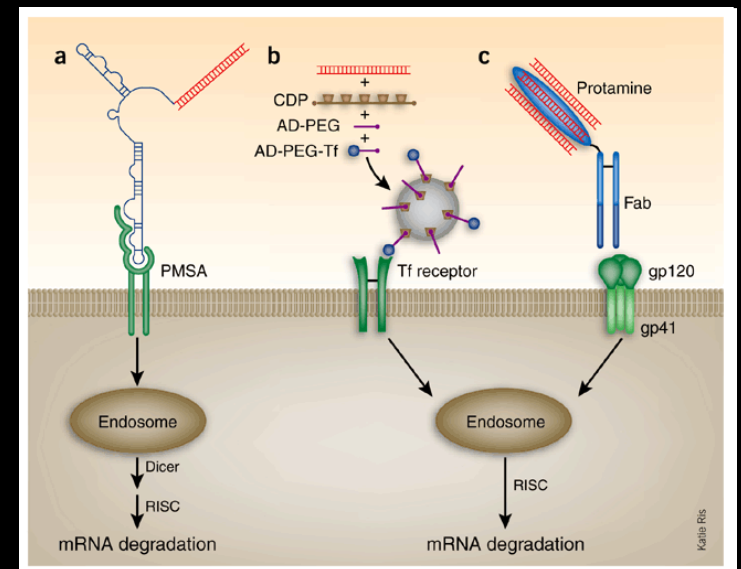
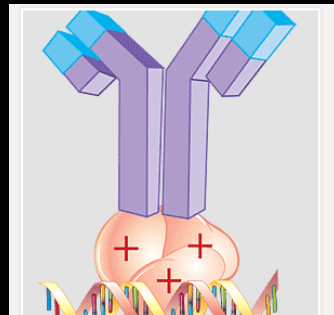
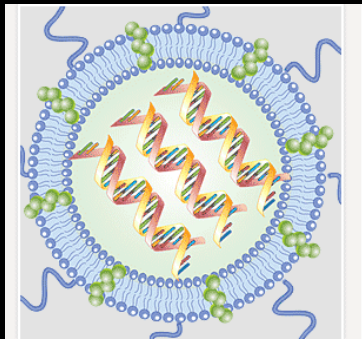
# siRNAs

- Synthetic small duplex RNAs (19-27 bp) with dinucleotide 3' overhang
- Chemical modifications
  - Improve serum stability
  - Minimize immunostimulation
  - Reduce off-target gene silencing (strand & seed)
  - Moieties for monitoring biodistribution
- Processed by Dicer and RISC (saturation concerns)
- Transient silencing following single-dose (duration)
- Readily dosable and can terminate Rx if adverse events
- Numerous algorithmic design tools on-line



# siRNA Delivery Hurdle

- Cationic liposomes or polymers
- Cholesterol-conjugates
- Peptide- and antibody-conjugates
- Aptamer-conjugates
- Various routes (direct injection or infusion, intravenous, inhalation)



# Some Promising siRNA Results to Date

- Silencing by siRNA infusion into mouse brain

Thakker et al. (2004) *PNAS*

- Nasally administered siRNA inhibit respiratory virus infection in mice

Bitko et al. (2005) *Nat Med*

- Lipoplexed siRNA prevented lethal infection of HSV-2 in mice

Palliser et al. (2005) *Nature*

- siRNA treatment improves macular degeneration (caveat)

Kleinman et al. (2008) *Nature*

- Systemic delivery of lipoplexed siRNA in mouse and non-human primate silences ApoB in liver

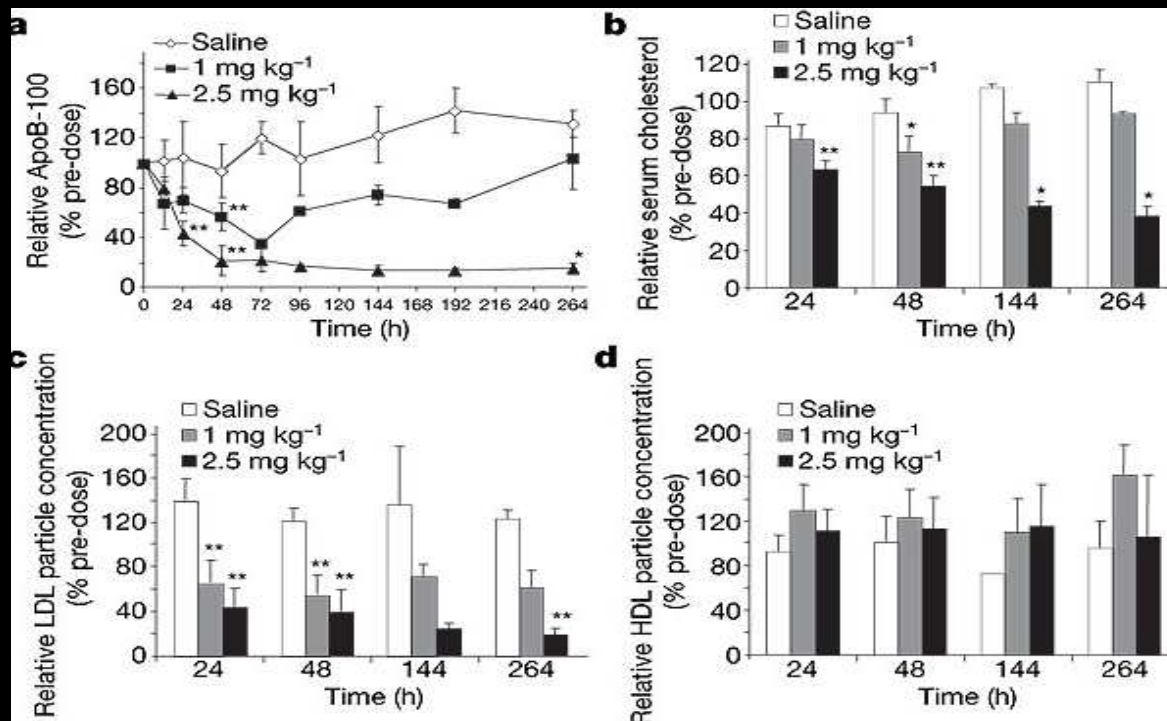
Zimmermann et al. (2006) *Nature*

- Systemic delivery of siRNA appears to work in humans

Davis et al. (2010) *Nature*

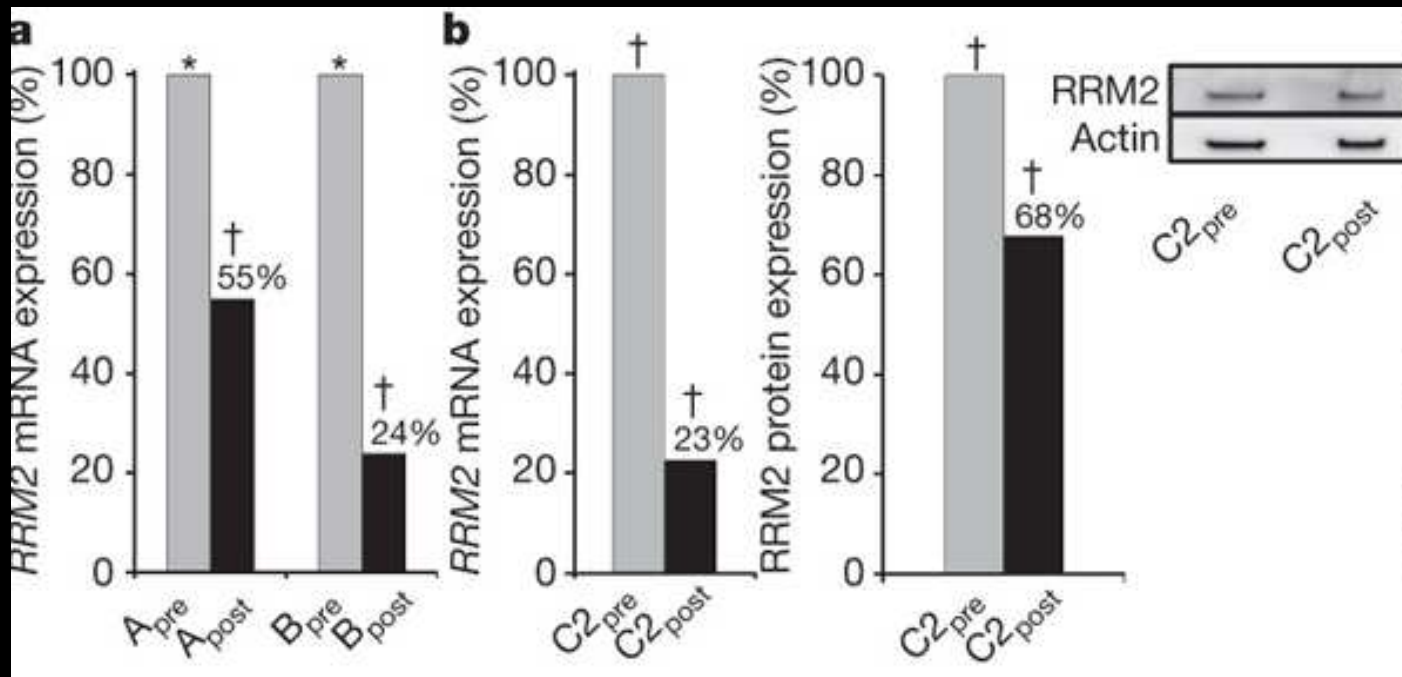
# Systemic siRNA in Non-Human Primate

- Cynomolgus monkeys were treated with single bolus i.v. injections of lipoplexed siApoB via the saphenous vein
- ApoB is expressed in the liver and is essential for secretion of VLDL and LDL, which are required for the transport and metabolism of cholesterol

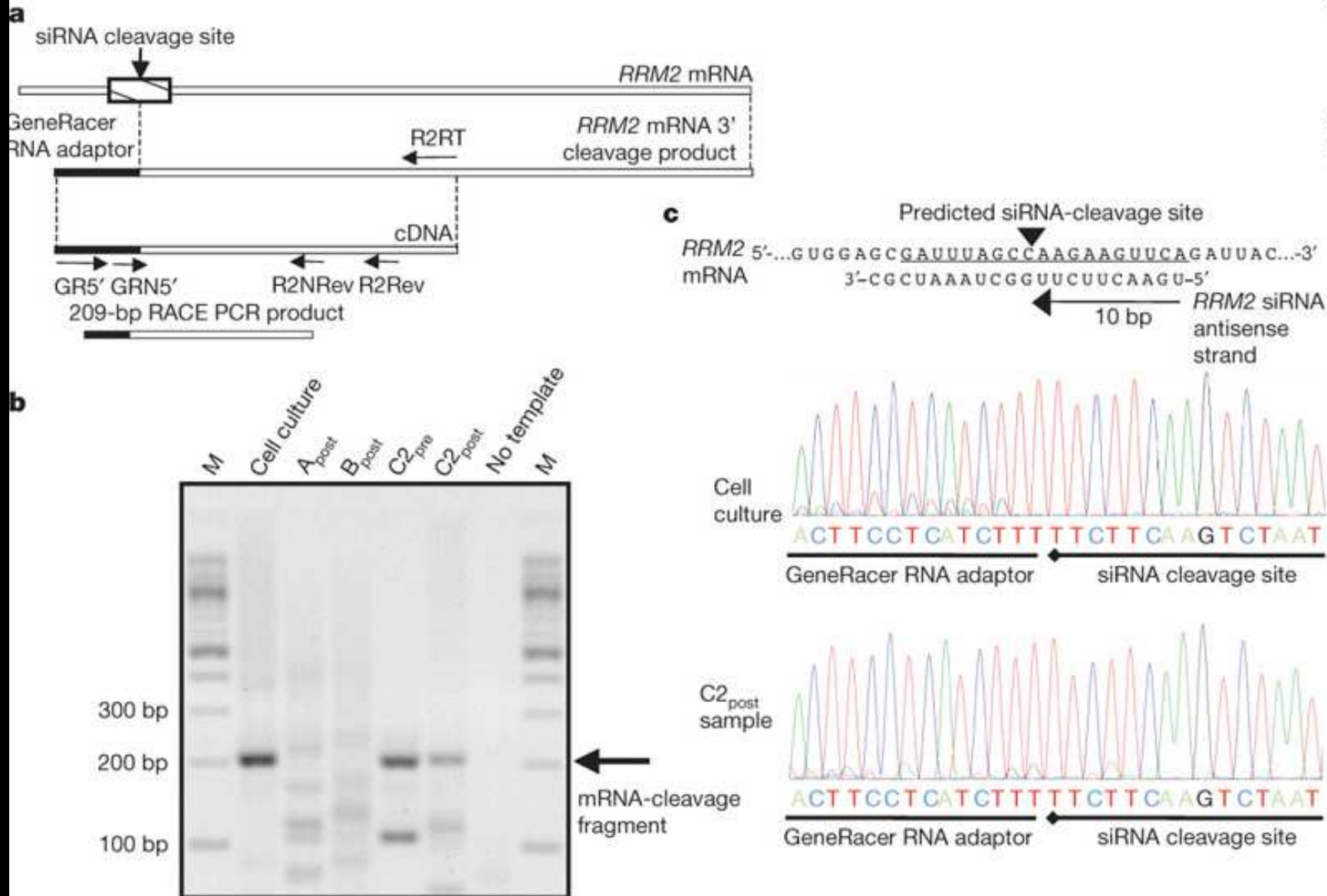


# Systemic siRNA in Human

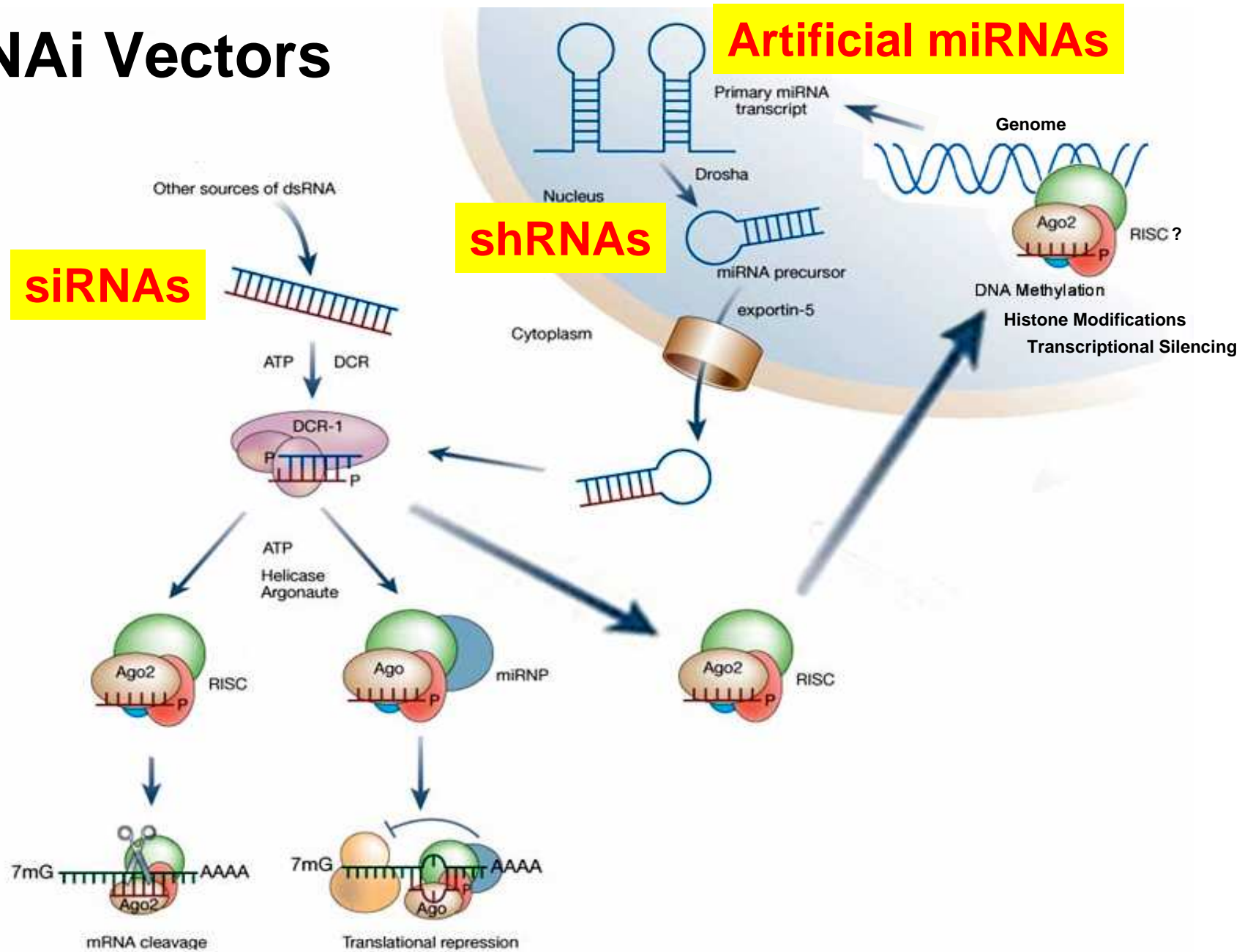
- siRNA clinical trials for macular degeneration, viral infection, cancer and skin disease are underway
- Systemic RNAi delivery remains a major hurdle
- Nanoparticles have recently shown promise in delivery of siRNAs to tumors via a systemic delivery route



# Systemic siRNA in Human



# RNAi Vectors

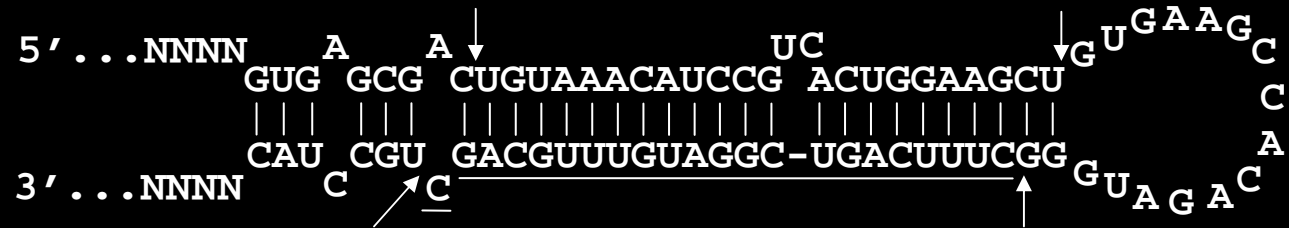


# RNAi Expression Vectors

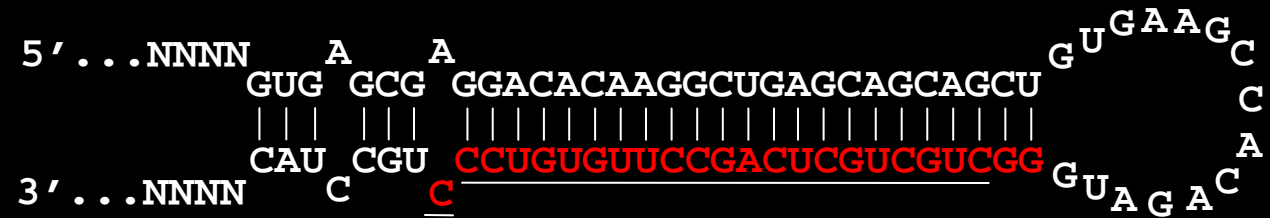
- Vector-based systems (plasmid, viral, etc.) are employed to express sense and antisense sequences to form duplex RNAs – most often in stem-loop formats
- Short-hairpin RNAs (shRNA)
  - Mimic pre-miRNAs (stem-loop with 3' overhang)
  - Substrate for Exportin-5, Dicer and RISC
  - Classically expressed from Pol-III promoters (e.g. U6 or H1) since start and stop transcription sequences should be controlled
- Artificial miRNAs or sh-miRs
  - Mimic pri-miRNA (embed siRNA in naturally occurring pri-miRNAs)
  - Substrate for Drosha, Exportin-5, Dicer and RISC
  - Can be expressed from any Pol-II promoter since transcription start & stop sites can be relatively arbitrary
- Allow persistent expression and long-term gene silencing following a single treatment

# RNAi Stem-loop Designs

mir-30



Artificial  
miRNA



shRNA





# Therapeutic Studies in Mice

<i>Disease / Virus</i>	<i>Target</i>	<i>Delivery Paradigm</i>	<i>Results</i>
Hepatitis B Virus	HBVsAg	Plasmid-shRNA to liver	Inhibited viral replication
HIV	Tat or CCR5	Lentivirus-shRNA to hematopoietic progenitors	Resistance to viral challenge
Cancer	BRAF, EGFR, VEGF, etc.	shRNA expressing viruses	Reduction in tumor volume, improved survival, etc..
Spinocerebellar ataxia type 1	ataxin-1	AAV-shRNA to Cerebellum	Improved histopathology & motor function
Huntington's disease	huntingtin	AAV-shRNA to Striatum	Reduced htt inclusions, improved motor phenotypes
Amyotrophic Lateral Sclerosis	SOD1	Lentiviral-shRNA to lumbar spinal cord	Improved motor neuron survival and function, extended survival
Alzheimer's	BACE1 or APP	Lentiviral-shRNA or AAV-shRNA to hippocampus	Reduced AB-deposition and improved neuropathology and learning & memory

Many more published results and many more studies underway.

# RNAi Toxicity in Mice (Liver)

## Fatality in mice due to oversaturation of cellular microRNA/short hairpin RNA pathways

Dirk Grimm<sup>1</sup>, Konrad L. Streetz<sup>1†</sup>, Catherine L. Jopling<sup>2</sup>, Theresa A. Storm<sup>1</sup>, Kusum Pandey<sup>1</sup>, Corrine R. Davis<sup>3</sup>, Patricia Marion<sup>4</sup>, Felix Salazar<sup>4</sup> & Mark A. Kay<sup>1</sup>

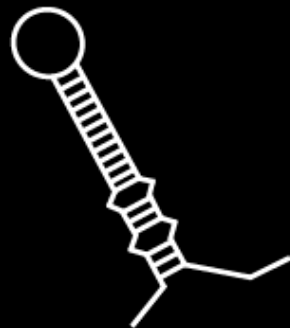
- Excessive doses of AAV8-shRNA vectors were delivered to the liver
- Mice receiving toxic shRNAs died within 30 days
- Toxicity correlated with shRNA expression levels and was, in part, due to saturation of RNAi processing machinery thus disrupting endogenous miRNA biogenesis and function. **Off-targeting?**

# shRNAs vs. Artificial miRNAs

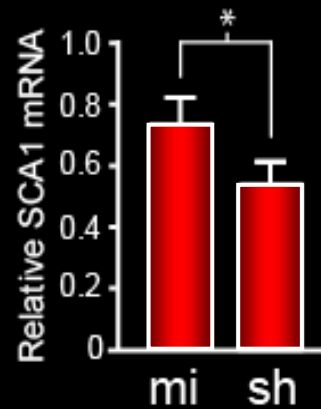
Comparison of silencing efficacies and RNAi processing



shRNA  
TOXIC



Artificial miRNA

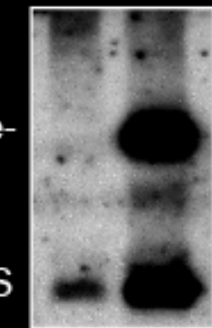


SCA1

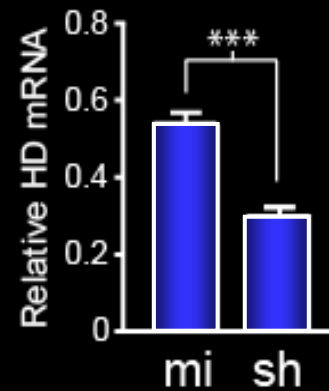
mi sh

Pre-

AS

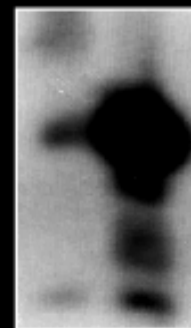


SCA1 RNAi

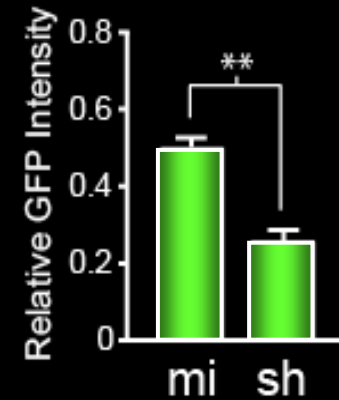


HD

mi sh

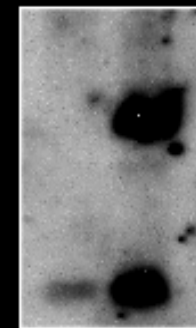


HD RNAi



GFP

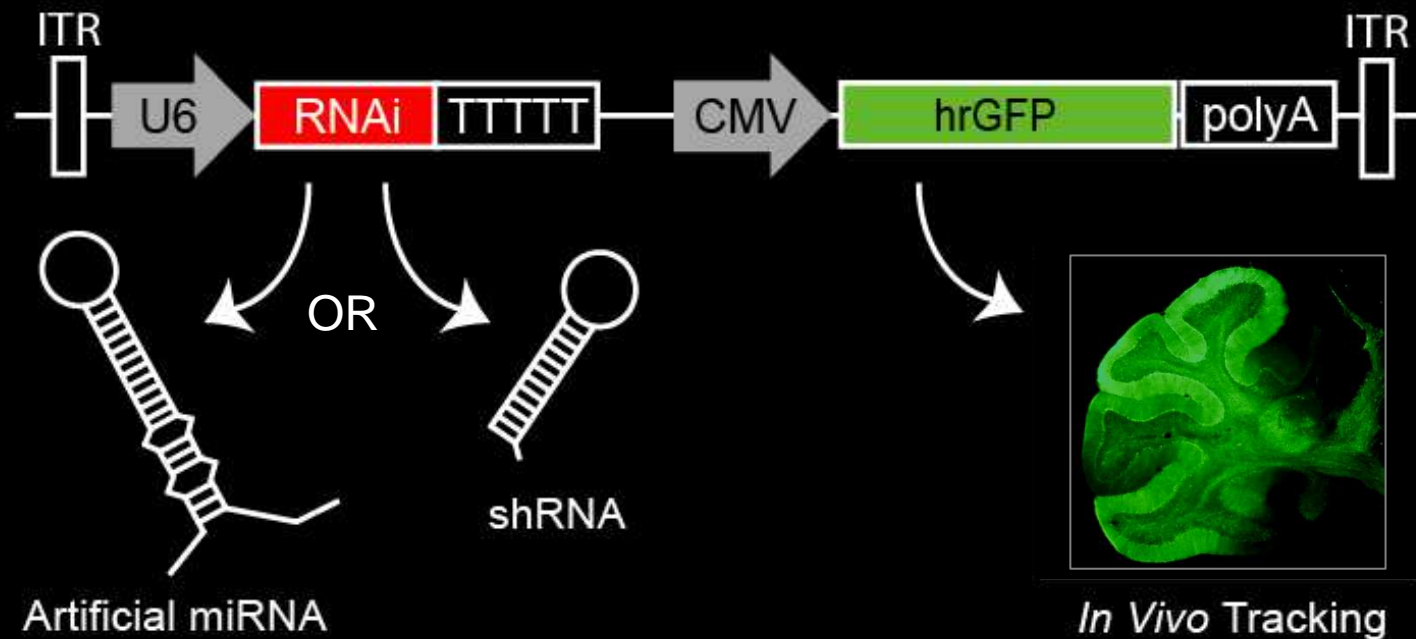
mi sh



GFP RNAi

# Evaluating RNAi Vector Safety and Efficacy in Mouse Brain

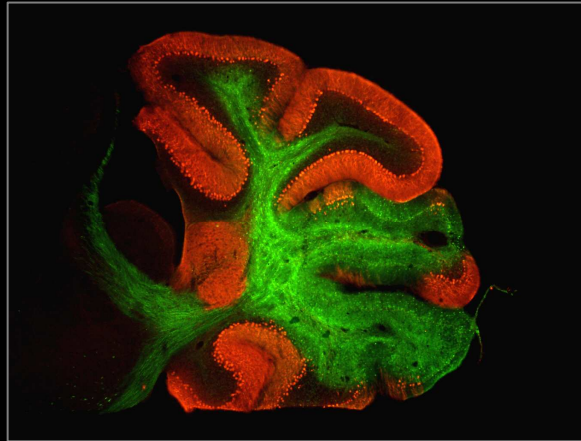
## Engineered AAV Genome



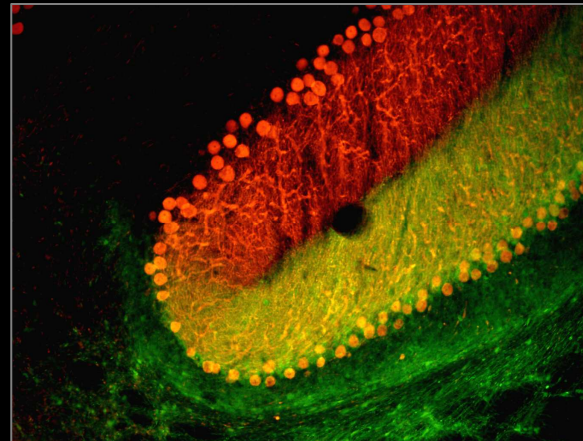
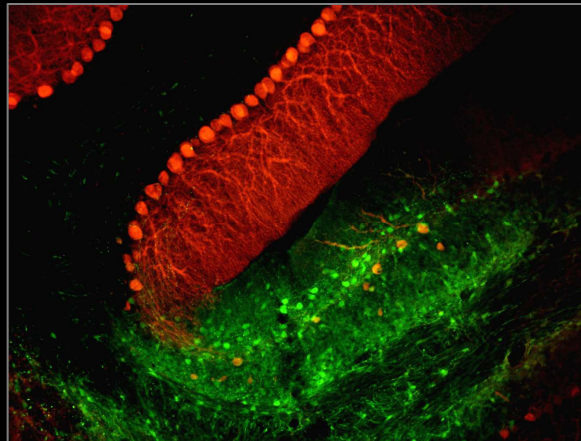
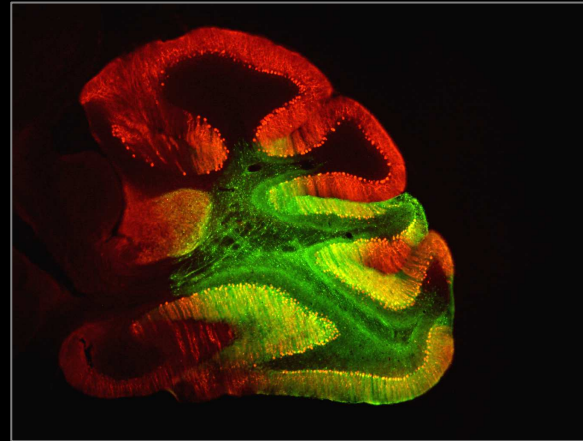
Clinically relevant vector system.

# RNAi Safety in Mouse Cerebellum

shSCA1

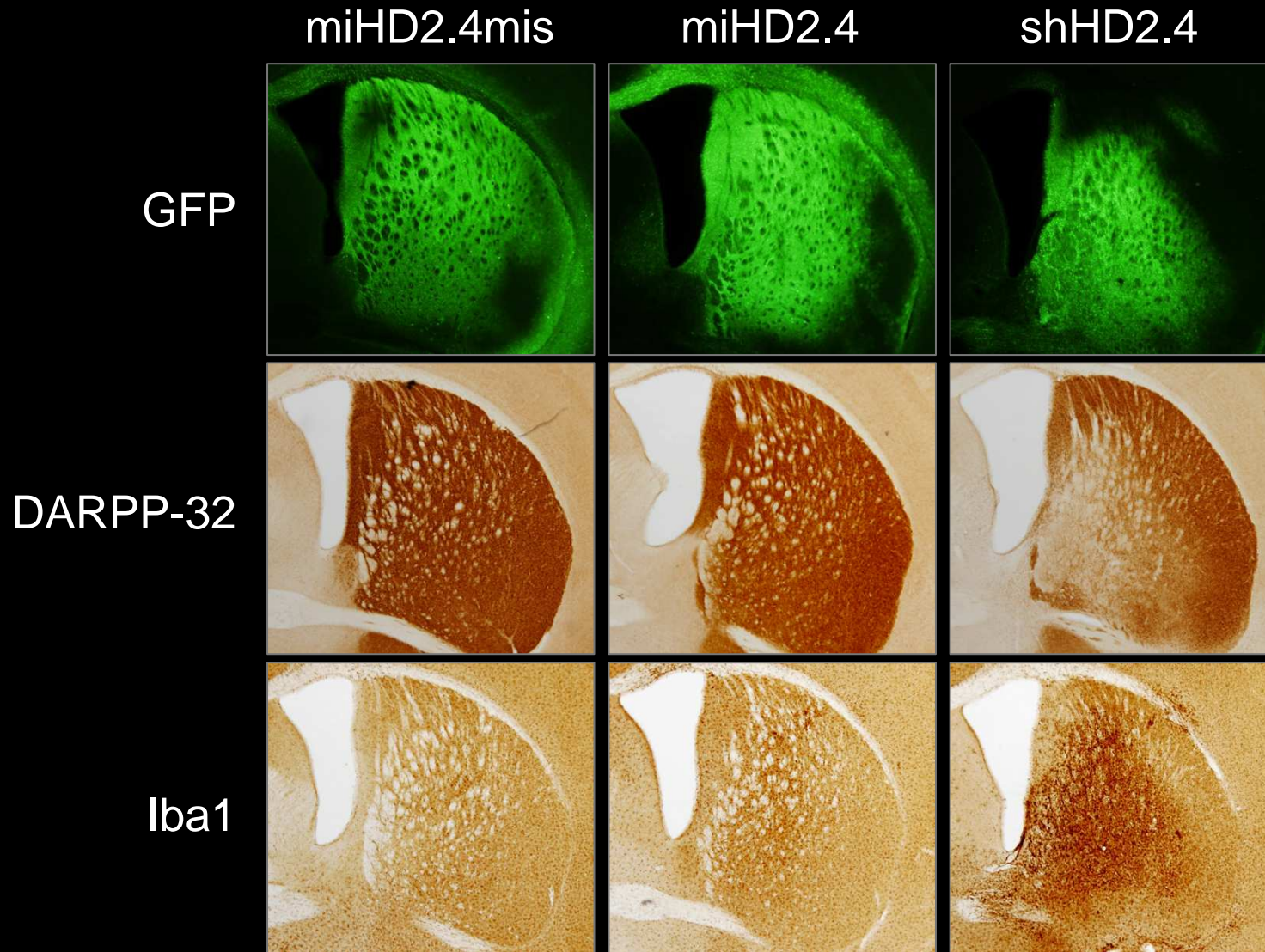


miSCA1

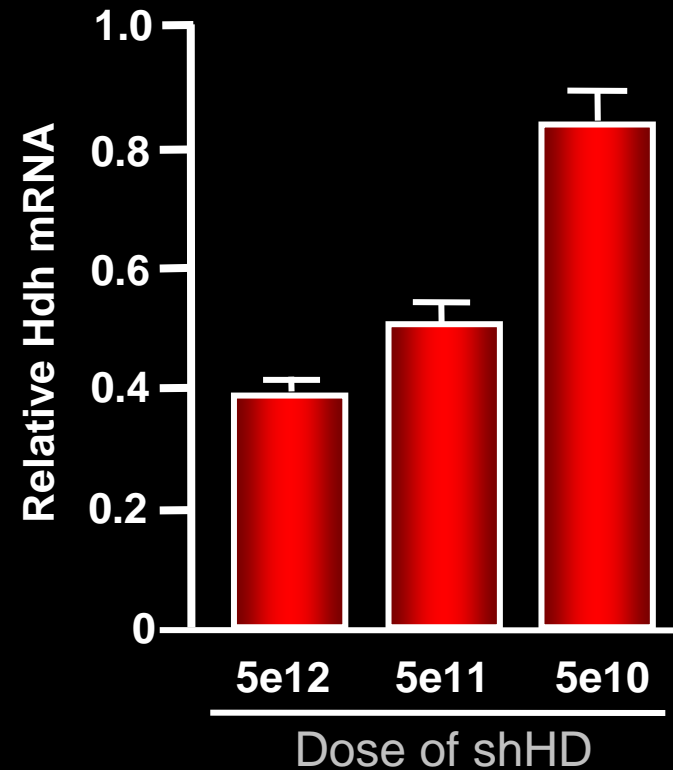
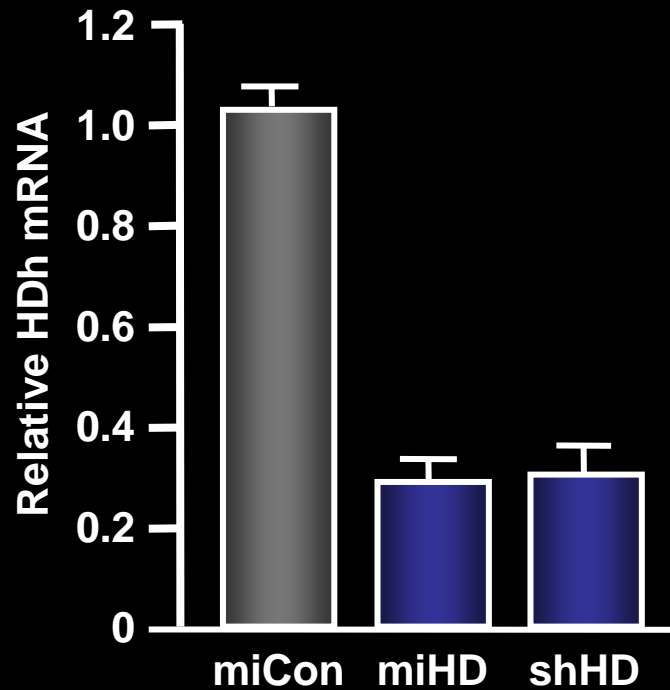


hrGFP-Calbindin

# Toxicity in Mouse Striata



# Silencing and Toxicity in Mouse Striata



HD2.4 AS



Lowering shRNA dose improved toxicity, but also led to ineffective target gene silencing.

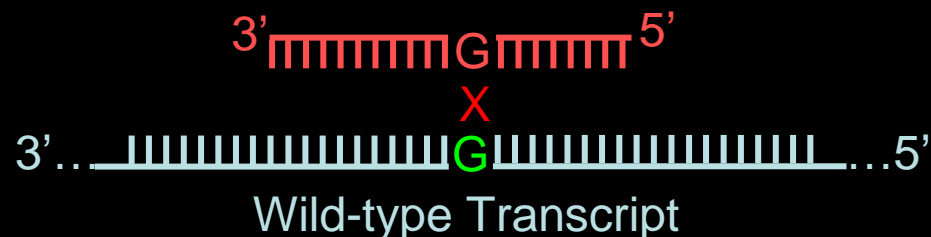
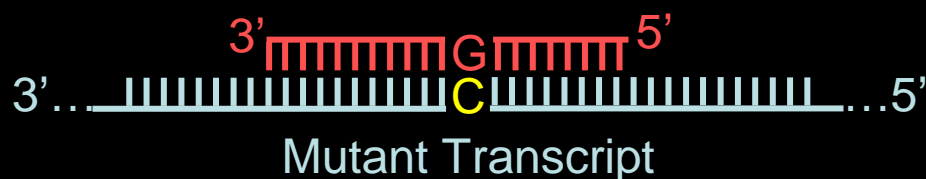
Why not deliver less shRNA virus?

# Importance of Dose Optimization

- Dose delivered (vector copies per cell)
- Promoter selection
- shRNA vs. artificial miRNA
- Optimization is dependent upon vector platform delivery modality, target tissue/cell population and therapeutic target

# Mutant Allele-Specific Silencing

- Relevant to dominant diseases
- If wild-type protein serves vital cellular function
- Target polymorphisms that discriminate wild-type and mutant transcripts
- Specificity can discriminate SNPs
- Has been investigated for a variety of diseases (AD, DYT1, HD, SCA3, SCA7)



# Future Directions & Considerations

- Delivery, delivery, delivery
- Employing safe and potent sequences will reduce concerns regarding saturation and off-targeting
- Regulated silencing for expression-based RNAi
- Clinical trials for every sequence?

# Questions?

*C. elegans*

Petunia



